

DOI: 10.14744/ejmo.2023.42324 EJMO 2023;7(2):95–102

**Review** 



# Epigenetic Code for Cell Fate During Development and Disease in Human

#### 💿 Selcen Celik Uzuner

Department of Molecular Biology and Genetics, Faculty of Science, Karadeniz Technical University, Trabzon, Türkiye

#### Abstract

Epigenetic reprogramming is the leading mechanism for cell differentiation in early development which gradually takes place upon zygote formation. This is governed by epigenetic modifications of genes involved in cell differentiation defined by Waddington's landscape. Somatic cells have specific gene expression profiles regulated by distinct epigenetic patterns. Therefore, they maintain their identity and specific gene profiles throughout lifetime. Although somatic cells can be induced into stem cell-like structures, the possible transformation of the cells can be associated with disruptions in cell identity leading to carcinogenesis. The epigenetic code for cell identity is the crucial player for maintaining stability and wellness of the cells during their lifespan. This review summarizes the epigenetic regulations involved in establishment of cellular fate and their abnormalities in cancer.

Keywords: Development, disease, epigenetics

Cite This Article: Celik Uzuner S. Epigenetic Code for Cell Fate During Development and Disease in Human. EJMO 2023;7(2):95–102.

## **Epigenetic Reprogramming**

Human begins his/her journey with a single cell called zygote. Zygote includes all the genetic information from ancestries provided by maternal and paternal pronuclei. Parental genetic background is then combined by pronuclei fusion followed by cell cleavage. Embryogenesis consists of sequential rounds of cell proliferation, and cell proliferation also occurs along with cell differentiation to end up with specified cells that further form embryonic layers and organs. Epigenetic reprogramming is the main mechanism that gives cells different identities even if they have the same DNA code. This is managed by a well-organised epigenetic machinery including DNA and histone modifications.<sup>[1]</sup>

The cloning of Dolly the Sheep was the groundbreaking discovery in the late 90's.<sup>[2,3]</sup> This discovery has opened the

doors to an approach for understanding of reprogramming of differentiated cells into a new embryo. This showed that something like a 'reverse evolution' is technically possible in developmental biology suggesting that not only stem cells are differentiated into somatic cells, but also somatic cells can be differentiated to stem cells. This indicates that the genome of a somatic cell has the potential to return to the first stage of its life as zygote. The method used for cloning of Dolly was "somatic cell nuclear transfer" (SCNT). Soon after the number of in vitro studies using SCNT method have focused on understanding the detailed principles of epigenetic reprogramming at gene and genome level. Although the Dolly was successfully cloned and lived for 7 years, the rate of live birth in such SCNT is low. One of the main reasons is the failure of epigenetic establishment derived by donor and/or transferred nucleus and the abnormalities in epigenetic modifications in cloned embryos.

Address for correspondence: Selcen Celik Uzuner, PhD. Department of Molecular Biology and Genetics, Faculty of Science,

Karadeniz Technical University, Trabzon, 61080, Türkiye

Phone: +90 462 377 20 32 E-mail: selcen.celik@ktu.edu.tr

Submitted Date: February 12, 2023 Revision Date: April 23, 2023 Accepted Date: April 28, 2023 Available Online Date: June 19, 2023 °Copyright 2023 by Eurasian Journal of Medicine and Oncology - Available online at www.ejmo.org

OPEN ACCESS This work is licensed under a Creative Commons Attribution-NonCommercial 4.0 International License.



<sup>[4,5]</sup> These epigenetic aberrations include i) errors in canonical and non-canonical genomic imprinting, ii) cell aging and iii) disruption in somatic cell-specific patterns of DNA and histone modifications.<sup>[3]</sup> Nevertheless, *in vitro* inducement of somatic cells into pluripotent stem cells provides explanations how a differentiated cell gains stem cell like characteristics suggesting that the cell fate is not necessarily unidirectional.

The sequential modification of specific genes either by DNA methylation or demethylation plays the critical role for the proper differentiation of cells.<sup>[6,7]</sup> Histone modifications also involve in the process of differentiation.<sup>[8,9]</sup> Therefore, the precise establishment of epigenetic patterns is a dynamic and inheritable mechanism in early development<sup>[11]</sup> and in adults it governs renewing some organs such liver.<sup>[10]</sup> Epigenetic pattern is a vital player from the very beginning of life, as cells gain their identity through epigenetic regulations. There was no life for multicellular organisms if there would no epigenetic reprogramming.

# Epigenetic Modifications: The Key Players in Epigenetic Reprogramming

Epigenetics investigates inheritable but dynamic and reversible chemical modifications occurring on DNA and histones that regulate gene expression. The principles of epigenetics can provide explanations for questions that cannot be explained by classical Mendelian Genetics.

Epigenetic modifications regulate cell differentiation processes to form tissues/organs from a zygote. Although all cells in a human body technically have the same DNA sequences, each cell type has its own epigenetic patterns.

Epigenetic modifications are mainly classified into two groups, DNA modifications and histone modifications (Fig. 1). DNA modifications on cytosine includes four sub-groups i) methylation (5meC) catalysed by DNMT (DNA methyltransferase) enzymes, ii) hydroxymethylation (5hmC), iii) formylation (5fC) and iv) carboxylation (5caC) by the oxida-

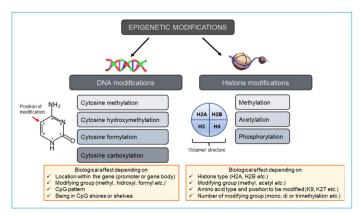


Figure 1. Main epigenetic modifications.

tion of 5meC with TET (Ten-eleven translocation) enzymes. <sup>[1]</sup> The common belief is that these are not the only epigenetic modifications on DNA, but more modifications are supposed to be identified in the near future. DNA methylation was previously thought to be associated only with gene inactivation, however today we know well that the function of DNA methylation depends on the modified location within the gene such as in promoters or in gene bodies. <sup>[11,12]</sup> Methylation at promoter regions is mainly associated with gene inactivation, however the methylation at gene bodies is associated with gene activation.<sup>[11-13]</sup> The other factor influencing the function of DNA methylation is the existence of methylation within the CpG repeats or not (such in shores, shelves and non-CpG regions) throughout the genome.<sup>[11,14]</sup> (Fig. 1, left panel).

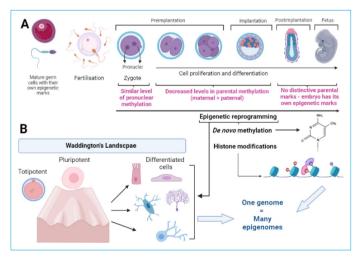
On the other hand, histones, the specific proteins composing chromatin structure, are the other critical player in epigenome. Histones can be modified by acetylation, methylation, phosphorylation, ubiquitination and SUMOylation.<sup>[15]</sup> Modifying group (methyl, acetyl etc.) is definitive for the biological function of modification. However, the type of modified histone (H2A, H2B, H3 and H4), the number of modifying groups (such di- or trimethylation) and the type of modified amino acid (such lysine or arginine) are also extremely important for the regulatory effects of histone modifications on gene expression profiles.<sup>[15]</sup> (Fig. 1, right panel). Histone modifications, in particular lysine acetylation, also specifically play critical roles in cellular activities including chromatin remodeling, cell cycle and splicing. <sup>[16]</sup> DNA and histone modifications do not independently function of each other and it is well-known that there is a dynamic crosstalk between the two machineries for the establishment, maintenance and regulation of gene expressions.<sup>[17,18]</sup> This fine-tuning of epigenetic modifications adds further complexity to understanding epigenetic reprogramming in normal and pathological processes.

Epigenetic modifications are dynamically regulated during early development. A wave of epigenetic programming occurs in the germ cell line to ensure that genomic imprinting is truly set. Genomic imprinting presents a model for monoallelic expression of some genes based on parent-oforigin. Genomic imprinting is regulated not only by DNA methylation but also histone modifications, particularly by H3K27me3.<sup>[19]</sup> Failure to maintain genomic imprinting is associated with a range of abnormalities in human.<sup>[20,21]</sup>

The paternal DNA methylation level is more than the maternal DNA methylation level at fertilisation. However in zygote, paternal and maternal methylation levels in pronuclei are found to be similar.<sup>[1]</sup> After fertilisation, the level of 5meC decreases by the oxidation mechanism catalysed by TET3 enzyme and accordingly the levels of the oxidised forms 5hmC, 5fC/5caC increase.<sup>[22]</sup> All cytosine modifications are then progressively erased until the blastula stage. TET1 and TET2 subsequently catalyse the increases in 5meC and 5hmC in blastula.<sup>[22]</sup> The understanding on reprogramming of other cytosine modifications (5hmC, 5fC and 5caC) remains more elusive than 5meC programming. After fertilisation, *de novo* methylation also occurs, and *de novo* methylation continues until implantation. After implantation of the embryo into the uterus, the embryo has its own epigenetic profiles including X chromosome inactivation<sup>[23]</sup> and tissue specific gene expression profiles.<sup>[24]</sup> This epigenome profile is established not only by DNA methylation and but also histone modifications.<sup>[25]</sup> (Fig. 2A)

One of the main objectives of epigenetic reprogramming is to form organs through the cell differentiation. This process is hypothetically explained by Waddington's landscape.<sup>[26]</sup> (Fig. 2B) This model suggests the progressive limitation of stem cell characteristics but inducement of tissue specific patterns. The cell fate is managed by epigenetic landscape. Each organ/cell has the same DNA code but the insulin protein, for instance, is only expressed in beta cells, a very specific pancreatic cell. Therefore, cells in a human body have the same genome, but hundreds/thousands of different epigenomes.

The epigenetic profiles of cells can be altered by environmental conditions. These conditions mainly refer to lifestyle such as eating habits,<sup>[27]</sup> educational attainment,<sup>[28]</sup> socioeconomic position,<sup>[29]</sup> psychiatric status<sup>[30,31]</sup> and smoking. <sup>[28]</sup> The field of "Environmental Epigenetics" investigates the epigenetic influence of human choices and external conditions. This is quite interesting that these epigenetic changes can be reversed after causal conditions, such as



**Figure 2.** Epigenetic changes in early development **(a)** and Waddington's. Landscape representing cellular differentiation **(b)** (adapted from<sup>[1]</sup>).

smoking, have ceased.<sup>[32]</sup> Reversible behaviour of epigenetic patterns on the genes offers a more dynamic and more manageable model for reprogramming than mutations.

# A Code in the Code: Cellular Memory, Cell Identity and Tissue-Specific Epigenomes

A human has a DNA (genome) that is identical in all cells, but there are numerous epigenomes that vary in different tissues (even within a tissue). This phenomenon suggests that there is another code in the DNA code, 'A code in the code', governed by epigenetic regulations. 'A code in the code' hypothesis defines the specific epigenetic profile for maintenance of cell identity, and somatic cells are therefore aware of which cell they are during their lifespan. The specific identity is established and characterised by specific gene expression and epigenomic profiles.<sup>[33–35]</sup> Somatic cells have an intelligence (a code) for managing themselves in terms of their identity. Somatic cells may be capable of "epigenetic reprogramming" even though they have already established their identity. But their ability to epigenetic reprogramming remains unclear.

Cellular memory is a phenomenon which is generally considered to be a property of immune cells or neurons. For instance, specialized immune cells called T and B cells (cytotoxic memory cells) can learn and memorize the information about antigens. Therefore, they reveal cytotoxic properties against these antigens/pathogens more easily. This pathogen knowledge can be inherited to the next generations to maintain the efficacy of the immune system cells. <sup>[36]</sup> But cellular memory of immunogenicity is not limited to specific immune cells, even fibroblasts in connective tissues have acquired a learned immunity against pathogenic microorganisms, and thus these cells have functions in the adaptive immune system.[37,38] Fibroblasts contain 1-10 receptors (TLRs, toll like receptors) that can recognize different microbial structures and activate immune system cells. <sup>[38]</sup> Fibroblasts are also involved in the repair of damaged cells by obtaining structural information from non-damaged cells in tissue damage.<sup>[39]</sup>

"Cellular memory" extensively represents the ability of cells to be aware of what type of cells they are throughout their lifespans. Cellular memory is also called as "transcriptional memory".<sup>[34,35]</sup> Each cell type has its own transcriptional memory represented by diverse profiles of gene expressions.<sup>[33]</sup> Memory for cell identity is shaped by epigenetic rearrangements in certain gene groups, and this memory is inherited to the next generations throughout the cell cycle. <sup>[40–42]</sup> Transcription factors are important for tissue-specific expression of genes by methylation-mediated specificity. <sup>[43]</sup> These transcription factors include for instance, HOXB13,

CDX1 and CDX2, which have affinity for binding methylated cytosines.<sup>[43]</sup> On the other hand, FoxA genes play a role for tissue-specific expression in hepatocytes and the methylation pattern of these genes is the key regulator for specific expression.[44] Interestingly, muscle cells have a memory of former physical activity, thus they have a high adaptation to retraining, and not surprisingly this memory is governed by epigenetic mechanisms, called 'epi-memory'.[45] Addition of neural-lineage transcription factor cocktail consisting of Ascl1, Brn2 and Myt1l converts fibroblasts to neuron cells. <sup>[46]</sup> Another transcription factor, NeuroD1, is also eligible to transform microglial cells into the neurons by mediating the histone alterations.<sup>[47]</sup> These suggest that reactivation of transcription factors by epigenetic regulations may alter cell fate, and somatic cells are able to be reprogrammed in vitro. But their capacity to reverse in vitro epigenetic manipulations remains unclear.

The pattern of DNA methylation is the key player to maintain tissue-specific expression in different types of cells. Lineage specific methylation governs the specification of the cells that further forms tissues and/or organs. This is also a defining factor to sub-specify cells derived in the same tissues. For instance immune system cells have distinct methylome patterns, and these patterns are associated with gene expression profiles. [48,49] The cells functioning in adaptive or innate immune response reveals different methylome marks.<sup>[49]</sup> Integrative analyses of omics data can provide details for understanding the cell-type specific characteristics, and this approach will further suggest predictive models for complex diseases.[48] Interestingly, DNA methylation is also associated with alternative splicing in a tissue-specific manner and this association may also be associated with abnormalities in diseases.<sup>[50]</sup>

#### Epigenetic Dysregulation Involved in Diseases

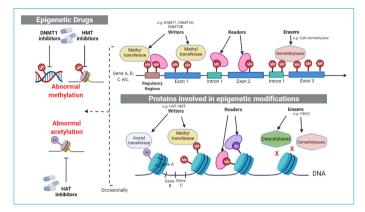
A line of evidence suggests that pathological processes involve abnormal patterns of epigenetic modifications.[51-53] Tumour suppressor genes and proto-oncogenes have been found to be mostly up- or downregulated by alterations in DNA methylation.<sup>[54–56]</sup> For instance a melanoma associated protein is upregulated by epigenetic mechanisms in an aggressive form of breast cancer.<sup>[51]</sup> Global hypomethylation is also commonly detected in different cancers,<sup>[57]</sup> but some cancers such as hepatocellular carcinoma, are significant with global hypermethylation.[58] Driver genes including tumour suppressor and proto-oncogenes are mostly found hypermethylated or hypomethylated in cancers, respectively.<sup>[59]</sup> Gene specific changes in DNA methylation are also highly informative for classification of cancers and estimation of the original tissue that cancer develops.<sup>[60]</sup> Neurodegeneration is also associated with the epigenetic changes

in some genes.<sup>[61]</sup> Some congenital diseases are associated with abnormalities in epigenetic code. For instance, some errors in genomic imprinting are related to Prader-Willi and Angelman syndromes.<sup>[20,21]</sup> DNMT1 depletion is lethal for human.<sup>[62]</sup> The lethal mutations in DNMT1 gene therefore resulted in abortion or foetus death in utero. Embryo cannot develop if DNMT1 enzyme is absent because cells can not differentiate and therefore not form organs. A cell does not value itself without any identity in multicellular organisms. Overexpression of DNMT1 gene is also associated with lethality in embryos or errors in genomic imprinting resulting in congenital abnormalities.<sup>[63]</sup> Increase in DNMT1 activity probably relates to misidentification of the cells.

## From Epigenetic Reprogramming to Epigenetic Targeting Drugs: A New Era in Therapy of Human Diseases

In some cases, the epigenetic changes that occur in a range of abnormalities such as cancer<sup>[64]</sup> and neuropsychiatric diseases<sup>[65]</sup> can be reversed by epigenetic drugs. Epigenetic targeting drugs, also known as 'epi-drugs', have been rather a new trend in disease treatments. These drugs, such 5'-aza-2'-deoxycytidine (Decitabine), have been widely used in cancer, and new drug candidates are in demand to discover more specific and effective drugs with minimum side-effects.[66-68] Low dose Decitabine is not cytotoxic for normal peripheral blood cells,<sup>[69]</sup> but are particularly cytotoxic to natural killer cells.<sup>[70]</sup> The main interest in epigenetic-based alterations of the genome has focused on carcinogenesis but cancer is not the only disease that is associated with epigenetic abnormalities. The diseases related to errors in genomic imprinting highlight the importance of correct establishment of epigenetic marks in the genome and there are epigenetic therapy approaches being considered for the treatment of genomic imprinting abnormalities.<sup>[71]</sup> It is important to maintain the paternalbased expression of alleles and this mechanism is governed by DNA methylation in the imprinted genes.

There are four main groups of proteins that regulate epigenetic modifications on DNA and histones; 1) writers, 2) erasers, 3) modulator proteins, and 4) mediator proteins<sup>[72]</sup> (Fig. 3). Writer proteins are the enzymes, such as DNMTs that add methyl groups to DNA, histone acetyltransferase (HATs) and histone methyltransferase (HMTs) enzymes that add acetyl and methyl groups to histones, respectively. Eraser proteins are the enzymes such as histone deacetylases (HDACs) removing the acetyl group from histones and CpG demethylase or TET1 removing the methyl group from DNA. Writer and eraser proteins are also called "Epigenetic Modifying Proteins". Modulatory proteins are the



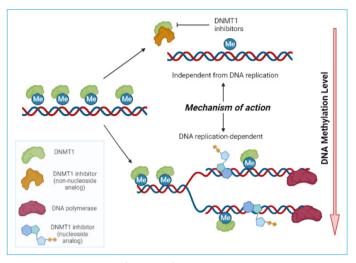
**Figure 3.** Proteins involved in epigenetic modifications and epigenetic targeting drugs (ETDs).

proteins that directly alter the epigenome through DNA methylation, histone modification, or structural changes of chromatin. For example, chromatin-modifying and re-modelling proteins are the modulator proteins. Mediator proteins are downstream targets of epigenetic modifications, and pluripotency factors such as OCT4, NANOG and SOX2.<sup>[73]</sup> These proteins collectively play role for epigenetic reprogramming in the cells.

Epigenetic drugs are mainly classified into two groups: i) DNA methylation inhibitors and ii) histone modification inhibitors (Fig. 3). A large group of these drugs involves enzyme inhibitors targeting DNMT1, HDAC or HAT enzymes to reverse the epigenetic marks.<sup>[74]</sup> DNMT1 inhibitors, which will be used in this project, include nucleoside or non-nucleoside analogues (Fig. 4). Nucleoside analogues, such 5'-azacytidine and 5'-deoxy-2'-azacytidine mimic the structure of cytidine, integrating into newly replicated DNA instead of methylated cytosine. These induce a gradual decrease in DNA methylation by each DNA replication. But non-nucleoside analogues act directly as inhibitors of DNMT1 enzyme to lower DNA methylation levels (Fig. 4). Although some of the DNMT1 inhibitors are clinically approved and used for treatment of a range of diseases, particularly in some cancers,<sup>[75,76]</sup> the main handicap of these drugs is that they do not target the specific sites in the genome, leading to undesired reprogramming in the epigenetic landscape. To deal with this limitation gene-specific approaches have been developed, including CRISPR-Cas9 and TET-based methods that can block DNA methylation and histone modifications in vitro on the specific sites.[77,78]

# The Missing Part of the Puzzle: Reprogramming Potential in Differentiated Cells

There are many studies mostly focused on how epigenetic reprogramming occurs in early development from stem cells to differentiated cells, and how induced reprogram-



**Figure 4.** Mechanism of action of DNMT1 inhibitors (nucleoside and non-nucleoside analogous).

ming can result in the transition of differentiated somatic cells to stem cells. The induced pluripotent stem cells (iP-SCs) created by somatic cell nuclear transfer are especially useful for therapeutic purposes. Somatic cells are supposed to maintain their cell identity by epigenetic tuning during normal processes. But this is not clear i) whether somatic cells can re-establish their ontological epigenetic patterns followed by epigenetic erasure by external agents (epigenetic targeting compounds), and ii) how they are back to their original state if they are able to re-build epigenetic profiles. The question is about whether/how differentiated human cells reprogramme their epigenetic motifs to maintain cellular identity after the loss of DNA methylation. Revealing the ontological response of normal cells against inducible changes in DNA methylation will encode the epigenetic mystery, and enlighten unknowns about disease progresses.

#### Disclosures

**Peer-review:** Externally peer-reviewed. **Conflict of Interest:** None declared.

#### References

- 1. Huntriss J. Epigenetic reprogramming in the embryo. Epigenetics Reprod Heal 2021:97–116.
- Niemann H. Epigenetic reprogramming in mammalian species after SCNT-based cloning. Theriogenology 2016;86:80– 90.
- Ogura A, Matoba S, Inoue K. 25th anniversary of cloning by somatic-cell nuclear transfer: Epigenetic abnormalities associated with somatic cell nuclear transfer. Reproduction 2021;162:F45–F58.
- 4. Dean W, Santos F, Stojkovic M, Zakhartchenko V, Walter J, Wolf E, et al. Conservation of methylation reprogramming in mam-

malian development: aberrant reprogramming in cloned embryos. Proc Natl Acad Sci U S A 2001;98:13734–8.

- Cao P, Li H, Zuo Y, Nashun B. Characterization of DNA methylation patterns and mining of epigenetic markers during genomic reprogramming in SCNT embryos. Front Cell Dev Biol 2020;8:570107.
- Fujii K, Tanaka S, Hasegawa T, Narazaki M, Kumanogoh A, Koseki H, et al. Tet DNA demethylase is required for plasma cell differentiation by controlling expression levels of IRF4. Int Immunol 2020;32:683–90.
- Izzo F, Lee SC, Poran A, Chaligne R, Gaiti F, Gross B, et al. DNA methylation disruption reshapes the hematopoietic differentiation landscape. Nat Genet. 2020;52:378–87.
- Gehre M, Bunina D, Sidoli S, Lübke MJ, Diaz N, Trovato M, et al. Lysine 4 of histone H3.3 is required for embryonic stem cell differentiation, histone enrichment at regulatory regions and transcription accuracy. Nat Genet 2020;52:273–282.
- Abe S, Nagatomo H, Sasaki H, Ishiuchi T. A histone H3.3K36M mutation in mice causes an imbalance of histone modifications and defects in chondrocyte differentiation. Epigenetics 2021;16:1123–34.
- 10. Aloia L. Epigenetic regulation of cell-fate changes that determine adult liver regeneration after injury. Front Cell Dev Biol 2021;9:346.
- 11. Lister R, Pelizzola M, Dowen RH, Hawkins RD, Hon G, Tonti-Filippini J, et al. Human DNA methylomes at base resolution show widespread epigenomic differences. Nature 2009;462:315–22
- Varley KE, Gertz J, Bowling KM, Parker SL, Reddy TE, Pauli-Behn F, et al. Dynamic DNA methylation across diverse human cell lines and tissues. Genome Res 2013;23:555–67.
- Yang X, Han H, De Carvalho DD, Lay FD, Jones PA, Liang G. Gene body methylation can alter gene expression and is a therapeutic target in cancer. Cancer Cell 2014;26:577–90.
- 14. Laurent L, Wong E, Li G, Huynh T, Tsirigos A, Ong CT, et al. Dynamic changes in the human methylome during differentiation. Genome Res 2010;20:320–31.
- 15. Bannister AJ, Kouzarides T. Regulation of chromatin by histone modifications. Cell Res 2011;21:381–95.
- Choudhary C, Kumar C, Gnad F, Nielsen ML, Rehman M, Walther TC, et al. Lysine acetylation targets protein complexes and co-regulates major cellular functions. Science 2009;325:834– 40.
- 17. Zhang B, Gu X, Han X, Gao Q, Liu J, Guo T, et al. Crosstalk between DNA methylation and histone acetylation triggers GDNF high transcription in glioblastoma cells. Clin Epigenetics 2020;12:47.
- Fu K, Bonora G, Pellegrini M. Interactions between core histone marks and DNA methyltransferases predict DNA methylation patterns observed in human cells and tissues. Epigenetics 2020;15:272–82.
- 19. Santini L, Halbritter F, Titz-Teixeira F, Suzuki T, Asami M, Ma X, et

al. Genomic imprinting in mouse blastocysts is predominantly associated with H3K27me3. Nat Commun 2021;12:3804.

- 20. Chang S, Wang Y, Xin Y, Wang S, Luo Y, Wang L, et al. DNA methylation abnormalities of imprinted genes in congenital heart disease: a pilot study. BMC Med Genomics 2021;14:4.
- 21. Elbracht M, Mackay D, Begemann M, Kagan KO, Eggermann T. Disturbed genomic imprinting and its relevance for human reproduction: Causes and clinical consequences. Hum Reprod Update 2020;26:197–213.
- 22. Pastor WA, Aravind L, Rao A. TETonic shift: Biological roles of TET proteins in DNA demethylation and transcription. Nat Rev Mol Cell Biol 2013;14:341–56.
- 23. Vallot C, Ouimette JF, Rougeulle C. Establishment of X chromosome inactivation and epigenomic features of the inactive X depend on cellular contexts. BioEssays 2016;38:869–80.
- Hasegawa Y, Taylor D, Ovchinnikov DA, Wolvetang EJ, de Torrenté L, Mar JC. Variability of gene expression identifies transcriptional regulators of early human embryonic development. PLOS Genet 2015;11:e1005428.
- 25. Burton A, Brochard V, Galan C, Ruiz-Morales ER, Rovira Q, Terrones DR, et al. Heterochromatin establishment during early mammalian development is regulated by pericentromeric RNA and characterized by non-repressive H3K9me3. Nat Cell Biol 2020;22:767–78.
- 26. Waddington C. How do cells differentiate? Sci Am 1953;189:108–17.
- 27. Shock T, Badang L, Ferguson B, Martinez-Guryn K. The interplay between diet, gut microbes, and host epigenetics in health and disease. J Nutr Biochem 2021;95:108631.
- 28. Gomez-Verjan JC, Esparza-Aguilar M, Martín-Martín V, Salazar-Perez C, Cadena-Trejo C, Gutierrez-Robledo LM, et al. Years of schooling could reduce epigenetic aging: a study of a Mexican cohort. Genes (Basel) 2021;12:1408.
- 29. Fiorito G, McCrory C, Robinson O, Carmeli C, Ochoa-Rosales C, Zhang Y, et al; BIOS Consortium; Lifepath consortium. Socioeconomic position, lifestyle habits and biomarkers of epigenetic aging: a multi-cohort analysis. Aging (Albany NY) 2019;11:2045–70.
- Perna L, Zhang Y, Matias-Garcia PR, Ladwig KH, Wiechmann T, Wild B, et al. Subjective mental health, incidence of depressive symptoms in later life, and the role of epigenetics: results from two longitudinal cohort studies. Transl Psychiatry 2020;10:323.
- Venditti S, Verdone L, Reale A, Vetriani V, Caserta M, Zampieri M. Molecules of silence: effects of meditation on gene expression and epigenetics. Front Psychol 2020;11:1767.
- 32. Keshawarz A, Joehanes R, Guan W, Huan T, DeMeo DL, Grove ML, et al. Longitudinal change in blood DNA epigenetic signature after smoking cessation. Epigenetics 2022;17:1098–109.
- 33. Lundberg E, Fagerberg L, Klevebring D, Matic I, Geiger T, Cox J, et al. Defining the transcriptome and proteome in three func-

tionally different human cell lines. Mol Syst Biol 2010;6:450.

- Kingston RE, Tamkun JW. Transcriptional regulation by trithorax-group proteins. Cold Spring Harb Perspect Biol 2014;6:a019349.
- 35. Francis NJ, Kingston RE. Mechanisms of transcriptional memory. Nat Rev Mol Cell Biol 2001;2:409–21.
- 36. Lefrançois L, Obar JJ. Once a killer, always a killer: From cytotoxic T cell to memory cell. Immunol Rev 2010;235:206–18.
- 37. Kaufman J, Graf BA, Leung EC, Pollock SJ, Koumas L, Reddy SY, et al. Fibroblasts as sentinel cells: role of the CDcd40-CDcd40 ligand system in fibroblast activation and lung inflammation and fibrosis. Chest 2001;120:53–5.
- 38. Hamada A, Torre C, Drancourt M, Ghigo E. Trained immunity carried by non-immune cells. Front Microbiol 2019;9:3225.
- Bustos-Arriaga J, García-Machorro J, León-Juárez M, García-Cordero J, Santos-Argumedo L, Flores-Romo L, et al. Activation of the innate immune response against DENV in normal non-transformed human fibroblasts. PLoS Negl Trop Dis 2011;5:e1420.
- Kim M, Costello J. DNA methylation: An epigenetic mark of cellular memory. Exp Mol Med 2017;49:e322.
- 41. Henikoff S, Greally JM. Epigenetics, cellular memory and gene regulation. Curr Biol 2016;26:R644–8.
- 42. Ringrose L, Paro R. Epigenetic regulation of cellular memory by the polycomb and trithorax group proteins. Annu Rev Genet 2004;38:413–43.
- 43. Yin Y, Morgunova E, Jolma A, Kaasinen E, Sahu B, Khund-Sayeed S, et al. Impact of cytosine methylation on DNA binding specificities of human transcription factors. Science 2017;356:eaaj2239.
- 44. Reizel Y, Morgan A, Gao L, Schug J, Mukherjee S, García MF, et al. FoxA-dependent demethylation of DNA initiates epigenetic memory of cellular identity. Dev Cell 2021;56:602–12.e4.
- 45. Wen Y, Dungan CM, Mobley CB, Valentino T, von Walden F, Murach KA. Nucleus type-specific DNA methylomics reveals epigenetic "memory" of prior adaptation in skeletal muscle. Function (Oxf) 2021;2:zqab038.
- 46. Vierbuchen T, Ostermeier A, Pang ZP, Kokubu Y, Südhof TC, Wernig M. Direct conversion of fibroblasts to functional neurons by defined factors. Nature 2010;463:1035–41.
- 47. Matsuda T, Irie T, Katsurabayashi S, Hayashi Y, Nagai T, Hamazaki N, et al. Pioneer factor NeuroD1 rearranges transcriptional and epigenetic profiles to execute microglia-neuron conversion. Neuron 2019;101:472–85.e7.
- 48. Chen L, Ge B, Casale FP, Vasquez L, Kwan T, Garrido-Martín D, et al. Genetic drivers of epigenetic and transcriptional variation in human immune cells. Cell 2016;167:1398–414.e24.
- 49. Roy R, Ramamoorthy S, Shapiro BD, Kaileh M, Hernandez D, Sarantopoulou D, et al. DNA methylation signatures reveal that distinct combinations of transcription factors specify human immune cell epigenetic identity. Immunity 2021;54:2465–80.e5.

- 50. Gutierrez-Arcelus M, Ongen H, Lappalainen T, Montgomery SB, Buil A, Yurovsky A, et al. Tissue-specific effects of genetic and epigenetic variation on gene regulation and splicing. PLoS Genet 2015;11:e1004958.
- Oh C, Kim HR, Oh S, Ko JY, Kim Y, Kang K, et al. Epigenetic upregulation of MAGE-A isoforms promotes breast cancer cell aggressiveness. Cancers (Basel) 2021;13:3176.
- Berry K, Wang J, Lu QR. Epigenetic regulation of oligodendrocyte myelination in developmental disorders and neurodegenerative diseases. F1000Res 2020;9:F1000 Faculty Rev–105.
- 53. Ohkura N, Sakaguchi S. Transcriptional and epigenetic basis of Treg cell development and function: its genetic anomalies or variations in autoimmune diseases. Cell Res 2020;30:465– 74.
- 54. Wu X, Cheng YL, Matthen M, Yoon A, Schwartz GK, Bala S, et al. Down-regulation of the tumor suppressor miR-34a contributes to head and neck cancer by up-regulating the MET oncogene and modulating tumor immune evasion. J Exp Clin Cancer Res 2021;40:70.
- 55. Xing H, Wang P, Liu S, Jing S, Lin J, Yang J, et al. A global integrated analysis of UNC5C down-regulation in cancers: insights from mechanism and combined treatment strategy. Biomed Pharmacother 2021;138:111355.
- Tokay E. Epidermal growth factor mediates up-regulation of URGCP oncogene in human hepatoma cancer cells. Mol Biol 2021;55:618–23.
- 57. Zhang W, Klinkebiel D, Barger CJ, Pandey S, Guda C, Miller A, et al. Global DNA hypomethylation in epithelial ovarian cancer: passive demethylation and association with genomic instability. Cancers (Basel) 2020;12:764.
- 58. Cerapio JP, Marchio A, Cano L, López I, Fournié JJ, Régnault B, et al. Global DNA hypermethylation pattern and unique gene expression signature in liver cancer from patients with Indigenous American ancestry. Oncotarget 2021;12:475–92.
- 59. Pfeifer GP. Defining driver DNA methylation changes in human cancer. Int J Mol Sci 2018;19:1166.
- Danilova L, Wrangle J, Herman JG, Cope L. DNA-methylation for the detection and distinction of 19 human malignancies. Epigenetics 2022;17:191–201.
- 61. Smith AR, Smith RG, Burrage J, Troakes C, Al-Sarraj S, Kalaria RN, et al. A cross-brain regions study of ANK1 DNA methylation in different neurodegenerative diseases. Neurobiol Aging 2019;74:70–6.
- 62. O'Neill KM, Irwin RE, Mackin SJ, Thursby SJ, Thakur A, Bertens C, et al. Depletion of DNMT1 in differentiated human cells highlights key classes of sensitive genes and an interplay with polycomb repression. Epigenetics and Chromatin 2018;11:12.
- 63. Biniszkiewicz D, Gribnau J, Ramsahoye B, Gaudet F, Eggan K, Humpherys D, et al. Dnmt1 overexpression causes genomic hypermethylation, loss of imprinting, and embryonic lethality. Mol Cell Biol 2002;22:2124–35.

- 64. Alaterre E, Ovejero S, Herviou L, de Boussac H, Papadopoulos G, Kulis M, et al. Comprehensive characterization of the epigenetic landscape in Multiple Myeloma. Theranostics 2022;12:1715–29.
- 65. Peedicayil J. The potential role of epigenetic drugs in the treatment of anxiety disorders. Neuropsychiatr Dis Treat 2020;16:597–606.
- 66. Montalvo-Casimiro M, González-Barrios R, Meraz-Rodriguez MA, Juárez-González VT, Arriaga-Canon C, Herrera LA. Epidrug repurposing: discovering new faces of old acquaintances in cancer therapy. Front Oncol 2020;10:605386.
- 67. Megiorni F, Camero S, Pontecorvi P, Camicia L, Marampon F, Ceccarelli S, et al. OTX015 epi-drug exerts antitumor effects in ovarian cancer cells by blocking GNL3-mediated radioresistance mechanisms: cellular, molecular and computational evidence. Cancers (Basel) 2021;13:1519.
- 68. Shi F, Li Y, Han R, Fu A, Wang R, Nusbaum O, et al. Valerian and valeric acid inhibit growth of breast cancer cells possibly by mediating epigenetic modifications. Sci Rep 2021;11:2519
- Brodská B, Holoubek A, Otevřelová P, Kuželová K. Combined treatment with low concentrations of decitabine and SAHA causes cell death in leukemic cell lines but not in normal peripheral blood lymphocytes. Biomed Res Int 2013;2013:659254.
- 70. Li X, Zhang M, Cai S, Wu Y, You Y, Wang X, et al. Concentrationdependent decitabine effects on primary NK cells viability, phenotype, and function in the absence of obvious NK cells proliferation-original article. Front Pharmacol 2021;12:755662.

- 71. Papulino C, Chianese U, Nicoletti MM, Benedetti R, Altucci L. Preclinical and clinical epigenetic-based reconsideration of beckwith-wiedemann syndrome. Front Genet 2020;11:1112.
- 72. Lu W, Zhang R, Jiang H, Zhang H, Luo C. Computer-aided drug design in epigenetics. Front Chem 2018 ;6:57.
- 73. Feinberg AP, Koldobskiy MA, Göndör A. Epigenetic modulators, modifiers and mediators in cancer aetiology and progression. Nat Rev Genet 2016;17:284–99.
- 74. Mohammad HP, Barbash O, Creasy CL. Targeting epigenetic modifications in cancer therapy: erasing the roadmap to cancer. Nat Med 2019;25:403–18.
- 75. Cheng H, Zou Y, Shah CD, Fan N, Bhagat TD, Gucalp R, et al. First-in-human study of inhaled Azacitidine in patients with advanced non-small cell lung cancer. Lung Cancer 2021;154:99–104.
- 76. Huls G, Chitu DA, Havelange V, Jongen-Lavrencic M, van de Loosdrecht AA, Biemond BJ, et al; Dutch-Belgian Hemato-Oncology Cooperative Group (HOVON). Azacitidine maintenance after intensive chemotherapy improves DFS in older AML patients. Blood 2019;133:1457–64.
- 77. Sapozhnikov DM, Szyf M. Unraveling the functional role of DNA demethylation at specific promoters by targeted steric blockage of DNA methyltransferase with CRISPR/dCas9. Nat Commun 2021;12:5711.
- 78. Shoaib M, Chen Q, Shi X, Nair N, Prasanna C, Yang R, et al. Histone H4 lysine 20 mono-methylation directly facilitates chromatin openness and promotes transcription of housekeeping genes. Nat Commun 2021;12:4800.